

The *p*-phenylphenacyl ester prepared in the usual way⁴ was the derivative of lactic acid.

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Several Derivatives of Acetyl-*dl*-phenylalanine¹

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In a recent communication, MacAllister and Niemann³ reported for the first time the hydrolysis of a hydrazide, *viz.*, nicotinyl-*l*-tyrosylhydrazide, by the proteolytic enzyme chymotrypsin. We are prompted to submit the preparation of another hydrazide, plus a thio ester, both of which are selectively attacked by beef chymotrypsin.

Experimental

Acetyl-*dl*-phenylalanine Hydrazide (APH).—APH was derived from acetyl-*dl*-phenylalanine ethyl ester (APEE),^{4,5} prepared in turn by the condensation of acetic anhydride and *dl*-phenylalanine ethyl ester. APEE (1.5 g.) was dissolved in 5 ml. of absolute alcohol to which was added 0.62 ml. of 100% hydrazine hydrate. The solution was permitted to remain in a stoppered flask for 24 hours at room temperature, after which time 1.25 g. of the desired compound, m. p. 160.1–160.2° (uncor.), was precipitated out with a mixture of ether and petroleum ether. It was recrystallized from ethanol, from which it separated as needles.

Anal. Calcd. for C₁₁H₁₅O₂N₃: C, 59.70; H, 6.83; N, 18.99. Found: C, 59.64; H, 6.68; N, 18.79.

Unlike nicotinyl-*l*-tyrosylhydrazide, APH is hydrolyzed slowly by chymotrypsin under the conditions observed. After a 20-hr. incubation at 24.6° of an aqueous solution (pH 7.3) consisting of APH (0.1 M) and chymotrypsin (0.470 mg. nitrogen/ml.) in 0.05 M phosphate buffer, the extent of hydrolysis was found to be 24.2% by titration of the carboxyl groups liberated.⁶ This value is based on an effective substrate concentration of 0.05 M, *i. e.*, it is presumed the *d*-form is not hydrolyzed. Blanks showed complete stability of APH, in the absence of enzyme, for a period of at least one day.

Acetyl-*dl*-phenylalanine Thio Ethyl Ester (APTEE).—Acetyl-*dl*-phenylalanine (5 g., 0.024 mole), thoroughly dried in a vacuum desiccator for a week over phosphorus pentoxide, was suspended in about 30 ml. of acetyl chloride. Phosphorus pentachloride (5.1 g., 0.024 mole) was added. The flask was immediately stoppered tightly and placed in an ice-bath, whereupon the acetyl-*dl*-phenylalanine rapidly dissolved. After the solution had remained at room temperature for three hours, it was vacuum distilled to dryness with total exclusion of moisture. Anhydrous ether was quickly poured on the residue, and the solution was again vacuum distilled to remove acetyl chloride and phosphorus oxychloride. The crude acyl

halide was further evacuated by pump for half an hour. Then about 40 ml. of ethyl mercaptan was added rapidly. Immediately a vigorous ebullition developed, and the residue went into solution. By the next day, some white material had precipitated out. The entire mixture was taken to dryness. The residue was slightly yellow and very hygroscopic. Water was added, the resulting suspension was chilled, made alkaline with bicarbonate, and stirred for five minutes with ether. The mixture separated into two layers; the ether layer was removed and dried over magnesium sulfate. The ether was removed by vacuum distillation to give a residue which still appeared somewhat yellow and hygroscopic. It was dried in a vacuum desiccator for several days. The residue was then washed with petroleum ether several times to remove the colored hygroscopic impurity. The crude thio ester (yield 3.2 g. or 53%) was dissolved in boiling ethanol, the solution cooled, and diluted with water. On standing in the ice-box, white needles crystallized out, m. p. 92–93° (uncor.).

*Anal.*⁷ Calcd. for C₁₃H₁₇O₂NS: C, 62.13; H, 6.82; S, 12.76. Found: C, 62.05; H, 6.78; S, 12.89.

APTEE (0.03 M) and chymotrypsin⁸ (0.048 mg. nitrogen/ml.) were incubated at 25° and an initial pH of 7.6 in a 50% alcoholic solution containing 0.006 M phosphate buffer. Within one minute an intense odor of ethyl mercaptan developed, accompanied by a drop in pH due to liberated carboxyl groups. The presence of mercaptan was confirmed by a positive nitroprusside test, which was carried out in weakly basic solution to avoid hydrolysis of APTEE. No spontaneous hydrolysis of the substrate could be detected in the absence of enzyme.

(7) Microanalyses of APH and APTEE by F. Schwarzkopf, Elmhurst, L. I.

(8) Once crystallized chymotrypsin was used to study the catalyzed hydrolysis of both APH and APTEE.

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The Hydration of 2-Heptyne¹

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The hydration of an unsymmetrically situated non-terminal carbon-carbon triple bond has been studied previously in only three cases. Stearolic acid³ was reported to form 9-keto- and 10-keto-stearic acids, by treatment with sulfuric acid, in a ratio of 42.4:57.6. In the same way, 9-undecyanoic acid yielded 59% 9-keto and 41% 10-keto product; hydration with acetic acid and mercuric acetate altered the ratio considerably, giving the 9-keto and 10-keto isomers in a ratio of 46:54.⁴ 2-Pentyne (the only hydrocarbon reported) yielded 2- and 3-pentanones in nearly equal amounts by the sulfuric acid method.⁵

Since there had been previously described⁶ a simple catalytic method for triple bond hydrations in high yield, it was decided to apply this procedure to 2-heptyne. A mixture of the two ketones, 2-heptanone and 3-heptanone, was easily

(1) Paper LV on substituted acetylenes; previous paper, *THIS JOURNAL*, **72**, 3542 (1950).

(2) Rev. Conrad J. Pillar, O.S.B., St. Benedict's College, Atchison, Kansas.

(3) Robinson and Robinson, *J. Chem. Soc.*, 2204 (1926).

(4) Sherrill and Smith, *ibid.*, 1501 (1937).

(5) Mowat and Smith, *ibid.*, 19 (1938).

(6) Thomas, Campbell and Hennion, *THIS JOURNAL*, **60**, 718 (1938).

(1) From the M.Sc. (June, 1949) and Ph.D. (November, 1949) Theses of Vivian Goldenberg and Harry Goldenberg, respectively, of the Polytechnic Institute of Brooklyn, New York.

(2) National Institutes of Health Predoctoral Fellow, 1947–1949.

(3) R. V. MacAllister and C. Niemann, *THIS JOURNAL*, **71**, 3854 (1949).

(4) APEE, m. p. 68.0–68.6°, was synthesized in excellent yield and high purity according to the directions of E. Fischer (*Ber.*, **37**, 2495 (1904)) for the preparation of the related compound, chloroacetyl-*l*-tyrosine ethyl ester.

(5) E. Cherbuliez and P. Plattner (*Helv. Chim. Acta*, **12**, 324 (1929)) reported a m. p. of 68°.

(6) W. Grassmann and W. Heyde, *Z. physiol. Chem.*, **183**, 32, (1929).